The Transitional Age Brain "The Best of Times and the Worst of Times"



Winston W. Chung, мD^a, James J. Hudziak, мD^{b,*}

KEYWORDS

- Transitional age youth Brain development Transitional age brain Behavior
- Neuroscience

KEY POINTS

- Over the past 2 decades, there have been substantial developments in the understanding of brain development.
- Progress in neuroimaging has allowed us to better understand the nuances of the development of cortical, subcortical, and white matter structures.
- Modern neuroscience, genomics, and epigenomic studies allow us a lens through which to develop an understanding of transitional age youth (TAY) behavior from a neurodevelopmental perspective.
- Developing brain building health promotion and illness prevention approaches for TAY will likely yield reductions in morbidity and mortality, enhance individual life trajectories, and have a life-long impact.

INTRODUCTION

A great deal of attention has been paid to the so-called zero to 3 period of brain development. Although clearly an important focus of neuroscience and public health, there is emerging evidence that a second critical period of neurodevelopment exists, bracketed by the onset of the peripubertal process to the completion of cortical organization (roughly ages 13–25). This 'transitional age brain' (TAB) epoch is marked by an increase in risk for morbidity, mortality, drug and alcohol use/misuse, and the onset of persistent psychiatric and nonpsychiatric medical conditions.

The central hypothesis is that the TAB has fully matured risk-taking hardware because of early maturation of subcortical brain regions (amygdala, nucleus

* Corresponding author.

E-mail address: James.Huziak@med.uvm.edu

Child Adolesc Psychiatric Clin N Am 26 (2017) 157–175 http://dx.doi.org/10.1016/j.chc.2016.12.017 1056-4993/17/© 2017 Elsevier Inc. All rights reserved.

childpsych.theclinics.com

Disclosure Statement: The authors have nothing to disclose.

^a Vermont Center for Children, Youth, and Family, University of Vermont Medical Center, 1 South Prospect Street, Arnold 3, Burlington, Vermont 05401, USA; ^b University of Vermont College of Medicine and Medical Center, 1 South Prospect Street, Arnold 3, Burlington, Vermont 05401, USA

accumbens, etc) but does not yet have matured regulatory hardware (fully pruned prefrontal and related cortical regions). We maintain that adolescents (13–17 of age) and their TABs benefit from imposed external regulatory systems in the form of parents, family members, teachers, and coaches. Even with external control and expectations, transitional age youth (TAY) remain at very high risk for morbidity and mortality associated with suicide, substance use and misuse, psychiatric illness, and accidents. At the same time that TAY and their maturing brains need more external regulatory support and lower risk environments, they instead have easier access to alcohol and drugs, high-risk social activities, and loss of close parenting and supervision. In other words, these negative environmental factors are in play at a very vulnerable time of brain development, in which the regulatory regions of the brain are undergoing the critical process of maturation.

Herein, we discuss the neurodevelopmental processes (with a special emphasis on pruning) that place TAY at high risk to make impulsive, poorly regulated decisions. We present a description of the symphony of brain development (neurogenesis, synaptogenesis, myelination, and pruning) from fertilization to the end of the TAB period to set the stage for just why the TAB epoch is a critical period. Although we emphasize the potential negative consequences of the TAB epoch, we also want to acknowledge that the same features of adolescent and young adult brain development may be a strength that allows TAY and TAB to respond to psychosocial interventions or to changes in environmental context with improved trajectories into adulthood.

EARLY BRAIN DEVELOPMENT

With advances in both structural (MRI) and functional (fMRI) imaging techniques, along with creative experimental designs using fMRI, information about the development of the human brain has been rapidly expanding. Still, research in the field continues to rely on studies the using other mammalian species with the extrapolation of data to humans.¹

Human brain development begins during the third week of gestation and continues to about the middle of the second decade, when the components involved in executive function become fully formed. It is a process that is intricate and tightly controlled, yet allows for some flexibility to adapt to the idiosyncrasies of the environment. One theme that emerges is that the brain matures by becoming more interconnected and each region becoming more specialized. Another theme that emerges is the process of overproduction before the elimination of excessive cells and connections based on experience, ultimately resulting in an efficient and unique processor.

The process begins with gastrulation, with differentiation of neural progenitor cells, followed by formation of the neural tube, then the neocortex and synaptic pathways. Around gestational week 28, the number of neurons in the human brain is at its peak, a level 40% greater than in adults. Dendritic growth, arborization, and synaptogenesis begin to accelerate rapidly.^{2–9} The rate of synaptogenesis reaches its peak around gestational week 34, but the net decrease of synapses does not begin to decrease until the onset of puberty.^{2,7–10} At the same time, up to 50% of the neurons undergo cell death to begin the process of establishing definitive connections.¹¹

After birth, neurogenesis is largely complete, and a dynamic process of synaptogenesis, myelination, programmed cell death, and pruning ensues as intrinsic and extrinsic signals interact. In the initial critical period up to age 3, development is dominated by synaptogenesis and by age 2 to 3, a toddler has more synapses than an adult and peaks at a level nearly twice that of adults.^{12,13} Some networks connections are exuberant, and entire networks are sometimes formed that are not found in adults.^{14–16} This process is regulated by neurotrophic factors such as brain-derived neurotrophic factor (BDNF), which blocks apoptosis.^{17,18}

Gray and white matter develop in concert with sensorimotor areas developing first, and then progressing to spatial orientation and language, before concluding with association regions (frontal lobe). There is a profound temporal mismatch between early subcortical development (limbic areas including the nucleus accumbens, amygdala, and others) and cortical regulatory regions. This period of cortical organization or pruning takes place during this second critical period during the transitional age (13–25 years), when the final pruning and myelination takes place. This leads to improved executive function such as attention, concentration, impulse control, reasoning, planning, problem solving, and mood regulation.

BRAIN PATTERNING AND PLASTICITY

During the prenatal period, a basic structure of brain organization is developed. The core structures from the spinal cord to the neocortex and the major compartments within these structures are formed, and there is an initial partitioning of the neocortex into well-defined functional areas.^{19–23} This initial patterning is underdefined, malleable, and based largely on intrinsic signaling.²⁴ Beginning in the late prenatal period, brain development is exquisitely responsive to extrinsic signaling or experience. Significant alterations to the brain structure can occur depending on whether the environment is enriched or deprived. This plasticity allows the mammalian brain to adapt to its environment.

The early patterning of the brain is well illustrated through the development of the neocortex. Emx2 and Pax6 are 2 signaling molecules produced in opposite gradients along the anterior–posterior axis in the neocortical proliferative zone.²⁵ The concentration of Pax6 is greatest in the anterior and lateral regions, whereas the highest concentration of Emx2 is in the posterior and medial regions. It is the relative concentration of each of these signaling molecules that contribute to the early patterning.^{19,25,26} High concentration of Pax6 and low concentration of Pax6 and a high concentration of Emx2 toward the caudal end lead to the development of the visual cortex (v1). The somatosensory cortex emerges in between, where the concentrations of Pax6 and Emx2 confirm that it is indeed the concentration of 1 signaling molecule in conjunction with the other that produces the specific patterning. When Pax6 expression is blocked, visual areas enlarge and somatosensory and motor areas shrink. When Emx2 expression is blocked, the opposite is true.

Despite this early patterning, the structure and function of these areas remain highly malleable and subject to experience. Greenough and coworkers have coined the term "experience expectant processes" to explain the phenomenon of synaptic and neuronal exuberance and subsequent pruning. This process allows for adaptation to ubiquitous environmental conditions that are common to all species.²⁷ However, it also suggests that the environment is a necessary condition for the normal development of certain neurobehavioral functions. An example of this would be the seminal study conducted by Hubel and Weisel on the early postnatal development of the visual cortex.^{28,29} By limiting the visual input of 1 eye on a primate, the bands of the active eye in the ocular dominance column expanded into the area of the deprived eye. Similarly, in Bachevalier and Mishkin's study, infant monkeys who sustained early lesions to the inferior temporal cortex were able to regain most of their function with only minimal deficits compared with adults who were not able to regain their function.³⁰ It is also noteworthy that infant monkeys that sustained medial temporal lobe lesions were not able to regain their function, suggesting that neural plasticity also has its limits.

Studies of human brain development also highlight the importance of experience and the environment. For example, poverty has been shown to be associated with significant impacts on brain development. Hanson and colleagues³¹ have shown lower total gray matter volumes, frontal and parietal volumes, and decreased total gray matter trajectory in children from lower income families. Additionally, early life stressors, which include low socioeconomic status, but also neglect and abuse, led to smaller amygdala and hippocampal volumes.³² Decreases in cortical gray matter thickness, right hemispheric volume, and left and right anterior insula volume^{33,34} are associated with early childhood depression. Interestingly, a variation of parent-child interaction therapy developed by Luby and associates,³⁵ named parent-child interaction therapy-emotion development, seems to be helpful for early childhood depression. Connectivity among regions important for emotional regulation is also similarly affected. Increased early life stressors was associated with decreased and atypical connectivity between the amygdala and the prefrontal cortex.^{36–39} And, Luby reported that lower income-to-need ratio was associated with reduced negative connectivity between left hippocampus and amygdala and the right superior frontal gyrus in addition to connectivity of both the amygdala and hippocampus bilaterally.⁴⁰

Although depravation can lead to thinning, atypicality, and loss of function, an enriched environment can have the opposite effect. Early on, Hebb noted rats reared in a "home" environment outperformed rats reared in a laboratory environment.⁴¹ Later, Rosenzweig and his colleagues also noted difference in brain weight, and other physical and histologic differences between rats reared in enriched versus impoverished environments.^{42–50} More recently, Greenough and coworkers showed that rats reared in a complex environment (large groups and objects in a cage that was frequently cleaned) had 20% to 25% more dendrites per neuron, and in those animals that were fitted with a monocular occluders, the exuberance was noted only unilater-ally.^{51–55} Moreover, the glial cells were also affected, with greater numbers and complexity, as well as an enhanced capillary system.^{51,56} Similar effects have been observed in adult rats reared in complex environments.^{57,58} In human studies, practice of specific skills such as juggling or playing a musical instrument has led to structural and white matter changes.^{59–69}

GRAY AND WHITE MATTER MATURATION

Early MRI morphometry studies compared children and adults. In Jernigan and Tallal's seminal study, gray matter volumes were shown to be considerably larger in school-aged children than in young adults.^{70,71} And in Giedd's landmark study, he showed that the volume of the cortical gray matter follow an inverted U shape, with peaks in late childhood and a surge just before puberty and occurring about 1 to 2 years earlier in girls.^{72,73} In subsequent studies, Shaw and Raznahan have also reported a curvilinear growth pattern of cortical thickness, a component underlying cortical volume along with cortical surface area.^{74,75} However, other studies have not identified the "inverted U" shape but rather monotonic linear decreases in cortical thickness. Sowell, Muftuler, Koolschijn, Mutlu, and Ducharme and their colleagues all primarily found first-order linear declines with the youngest children in this study being around 5 years old.^{76–80} More important, Ducharme and associates⁷⁸ found postprocessing quality control procedures significantly impacted the complexity of the growth trajectories.

When no quality control procedures were implemented, there were more areas with quadratic and cubic trajectories. The biological underpinnings of the volumetric and cortical thickness changes are owing to arborization as well as continued glial cell maturation opposed by pruning of neuronal processes.¹¹ Changes in neuronal size, glial cell density, and vasculature could also contribute to cortical thickness changes.⁸¹

More recent structural MRIs allowed for more precise measurements of cortical thickness.^{82,83} These more detailed studies were able to show a modal pattern of cortical development, suggesting regional specificity in cortical thickness.⁸¹ Gogtay and coworkers⁸⁴ reported that phylogenetically older regions matured (thinned) earlier than newer regions. They also observed that the maturation process starts with lower order somatosensory and visual cortices before proceeding to multimodal and supramodal cortices.⁸⁴ The lateral prefrontal cortex and the temporal poles, regions processing motivation, goal-setting, and integration of emotion, are the last regions to mature.⁸⁴⁻⁸⁷ The maturation of various cortical regions has been correlated with performance measures. Maturation of the prefrontal cortex is related to cognitive intelligence, the anterior cingulate cortex is related to impulse control, the motor cortex is related to fine motor skills, and the left hemisphere area is related to increased language processing skills.^{88–91} Our group has shown that, in typically developing children followed across development, it is possible to relate specific cortical thickness regions in whole brain analyses that correlate with quantitatively different expressions of common traits such as aggression, anxiety, attention, emotional regulation, and externalizing problems.92-96

The subcortical gray matter also undergoes significant changes, although to a lesser degree compared with the neocortex.^{79,97–100} The caudate, which has traditionally been implicated in control of movement and muscle tone, and more recently in mediating higher order cognitive functions, had previously been shown in some but not all studies to also have a curvilinear developmental trajectory with peaks during the pre-adolescent to adolescent years.^{101,102} However, in a more recent study, Ostby and associates¹⁰⁰ analyzed multiple structures in the basal ganglia (caudate, putamen, and pallidum), and nucleus accumbens, and found a linear decrease over time. The hippocampus and the amygdala on the other hand, showed an increase in size with age.^{97–99,103,104}

Unlike gray matter, white matter volume begins to increase in the postnatal period and continues into middle adulthood.^{105–107} The development of diffusion tensor imaging has allowed the visualization of white matter by measuring proton diffusivity. As fiber tracts mature and myelination proceeds, diffusion or fractional anisotropy increases.¹⁰⁸ Although many studies found that white matter volume linearly increases over time, some studies have suggested it may have a curvilinear inverted U function, reaching its peak during the second decade.^{71,72,109,110} Also like gray matter, different tracts mature at different times. Tracts such as the corticospinal tract and corona radiata are mature by adolescence, but tracts responsible for executive function and emotional and behavioral control such as the internal capsule are still maturing in the adolescent. The continued maturation of these tracts likely is a major contributor to the improvements in modulation of adolescent behavior.^{111–113} The maturation of the white matter tracts has implications for other cognitive functions, such as intelligence, visuospatial skills, response inhibition, memory, reading skills, and language.¹¹⁴

The nuances in the development of various cortical and subcortical regions are noteworthy, but further investigations with a longitudinal design are needed to confirm the results.

TRANSITIONAL AGE BRAIN MISMATCH HYPOTHESIS

In almost every measurable domain, adolescence is a developmental period of strength and resilience. Compared with young children, adolescents are stronger, bigger, and faster, and are achieving maturational improvements in reaction time, reasoning abilities, immune function, and capacity to withstand cold, heat, injury, and physical stress. Yet, despite these robust maturational improvements in many domains, overall morbidity and mortality rates increase by 200% over the same interval of time.¹¹⁵ This doubling in rates of death and disability from the period of early school age into late adolescence/early adulthood is likely owing to difficulties in the control of behavior and emotion. This period is, therefore, marked for the high rates of accident suicide, violence, and health problems related to risky sexual behaviors.¹¹⁵ In addition, adolescence is the peak time of emergence for several types of mental illnesses, including anxiety disorders, bipolar disorder, depression, eating disorders, psychosis, and substance abuse. In the National Comorbidity Survey Replication Study, Kessler and colleagues¹¹⁶ found that 50% of most mental illnesses people experience emerge by age 14 and 75% start by age 24.

A simplistic explanation of the developmental mismatch can be understood through the emerging understanding of the development of 3 key regions of the brain: the nucleus accumbens, amygdala, and prefrontal cortices.

The nucleus accumbens is the home of motivation, passion and pleasure. Afferents from the ventral tegmental area, specifically the A9 and A10 nuclei travel over the mesolimbic dopamine pathway to stimulate the nucleus accumbens via dopaminergic neurons. Activity in the nucleus accumbens influences how much effort an individual will expend in pleasure and reward-seeking behavior. A developing nucleus accumbens is believed to contribute to the often observed tendency that some TAY prefer activities that require low effort yet produce high excitement. When mature, the mesolimbic dopaminergic pathway terminates in prefrontal and frontal cortical regions and is responsible for attentional control, concentration, and mood regulation. However, those frontal regions are not yet organized and the mismatch of drive without control is the essence of the TAB and central to the developmental mismatch theory.

The amygdala plays a key role in both emotional recognition and regulation. It increases in volume from late childhood until late adolescence, with a decelerating rate of growth after the age 16.¹¹⁷ Pathways into and out of the amygdala are responsible for integrating emotional reactions to both pleasurable and aversive experiences. It is hypothesized that, during the developmental mismatch epoch, the amygdala contributes to 2 behavioral effects: the tendency for adolescents to react to situations with "hot" emotions rather than more controlled and "cool" emotions and the propensity for youth to misread neutral or inquisitive facial expressions from other individuals as a sign of anger.⁷⁴ As the TAB matures, the functional connectivity of the amygdala changes dramatically and amygdala activity becomes more closely linked to activity in those prefrontal cortex regions involved in emotional regulation. This signals an end to the TAB period and the beginning of a more neurologically regulated period referred to as adulthood.¹¹⁸

Last, the prefrontal cortex is the key area of the brain involved in attention, emotional regulation, impulse control, cognitive flexibility, planning, and judgment. To do this, it processes complex information, requiring inputs from many brain regions. It allows a person to make decisions and select a course of action based on impulse inhibition, foreseeable consequences, and personal goals. It also happens to develop much later than the nucleus accumbens or the amygdala because cortical thinning or gray matter volume loss (maturation) occurs from the back of the brain to the front. In fact, the

prefrontal cortex is the last brain region to complete development. As a consequence, TAY rely much more on the emotional regions of their brain, such as the amygdala and other limbic regions, to guide their behavior compared with adults.^{74,119} In other words, TAY do not yet have the brain structures in place that allow the consistent use of the prefrontal cortex for its primary responsibilities of planning, judgment, cognitive flexibility, and impulse control. This mismatch in maturational timing is most exaggerated during the adolescent period, when the subcortical structures including the hormone fueled limbic system is already developed while the prefrontal cortex continue to mature.¹²⁰ Most recent studies seem to bear this out, indicating that risk-taking behaviors, sensation seeking, and heightened emotional reactivity are linked to the developmental mismatch in maturation of cortical and subcortical structures.¹²¹ The fact that puberty seems to be starting earlier means that the "mismatch years" are being extended.

As a result of this mismatch, TAY at times use poor judgment and take excessive risks. The relative delay in the maturation of frontal cortical areas as well as those networks connecting to them leaves adolescents vulnerable to performance impairments of higher order cognitive functions, such as cognitive control and response inhibition when stressed.^{122–125} TAY have increased activation of the ventral striatum to rewards and decreased activation during reward anticipation, which leaves them vulnerable to risk taking.^{126–130} Further, their amygdala are less activated in response to aversive outcomes while exhibiting social emotional bias, resulting in more risky behavior in the presence of others.^{131–135} As such, they more frequently make poor decisions and take excessive risk.⁸⁴

Similarly, the protracted development of the uncinated fasciculus provides insight into the difficulty TAY have in emotional regulation. The uncinated fasciculus is a fiber pathway composed of frontotemporal connections that include projections from orbitofrontal cortex to the anterior temporal cortex, as well as the amygdala.^{136,137} It has been shown to display age-related increases in microstructural integrity well into adulthood.¹³⁸ Although the amygdala has been implicated in mediating negative affective states such as fear and anxiety, hemodynamic activity within aspects of the orbitofrontal cortex has been associated with emotion regulatory processes.¹³⁹ The continued development of the orbitofrontal cortex and this pathway that links it to the amygdala is what allows for improved affective regulation as TAY grow into adulthood.

EMOTION REGULATION AND THE BRAIN

The neuropsychological processes important for emotion perception, allowing the generation of contextually appropriate, complex affective states, emotional experiences (feelings), and behaviors, include the identification of the emotional significance of an environmental stimulus, the production of an affective state and emotional behavior, and the regulation of the affective state and emotional behavior. Findings of neuroimaging studies indicate that specific neural regions may be important for more than one of these processes. The processes may be depend on the functioning of 2 main neural systems: a ventral system and a dorsal system. The ventral system, including the amygdala, insula, ventral striatum, and ventral regions of the anterior cingulate gyrus and prefrontal cortex, is important for the identification of the emotional significance of environmental stimuli and the production of affective states. It is additionally important for automatic regulation and mediation of autonomic responses to emotive stimuli and contexts accompanying the production of affective

states. The dorsal system, including the hippocampus and dorsal regions of the anterior cingulate gyrus and prefrontal cortex, regions where cognitive processes are integrated, is important for the performance of executive functions, including selective attention, planning, and effortful rather than automatic regulation of affective states.¹⁴⁰

The emotional circuitry of the brain is being shaped continuously by experiences that impinge on the nervous system during prenatal development and throughout life. This experience-induced plasticity has been documented in the brain in a variety of animal models and there is now substantial evidence on the effect of stressful and stimulating environments on the developing human brain and associated behavior. Most evidence is obtained by structural MRI studies of children that experienced adversity. Abused children are shown to have smaller orbitofrontal volumes and, the smaller the orbitofrontal volume in the abused sample, the more severe the social stress was reported.¹⁴¹ In another study, higher parental ratings of internalizing behavior and anxiety were correlated with a larger amygdala volume. Furthermore, children continuously exposed to maternal depressive symptoms from birth had significantly larger left and right amygdala than children with no such exposure.¹⁴² Such a developmental pattern in the amygdala has been suggested to occur in the autistic brain.^{143,144} Research suggests that some of these alterations in brain structure are caused by epigenetic regulations. For example, child abuse is associated with alterations in the epigenetic regulation of the glucocorticoid receptor extracted from the hippocampus of suicide victims with a history of child abuse compared with those with no abuse history along with controls. In the hippocampus, decreased levels of glucocorticoid receptor messenger RNA were observed.¹⁴⁵ Several pathologic conditions (eg, major depression, posttraumatic stress disorder) are associated with decreased density or volume of the hippocampus.^{146,147}

EPIGENETICS, GENE EXPRESSION, AND A PERIOD OF VULNERABILITY AND OPPORTUNITY

As discussed, epigenetic regulations can have powerful effects on the brain. The main factors that contribute to epigenetic changes in the adolescent brain are DNA methylation, histone modification, and microRNAs or noncoding RNAs. DNA methylation is generally associated with gene silencing as a methyl group binds to CpG islands blocking RNA polymerase, although recent evidence suggests that the effects of methylation could be more complicated.^{148–150} Histone modifications alter the accessibility of the transcription machinery to the DNA and include methylation, acetylation phosphorylation, and ubiquitination.¹⁵¹ And last, microRNAs are small noncoding RNAs that regulate posttranscriptional gene expression by affecting the translational efficiency of specific messenger RNA targets.^{152–154} Noncoding RNAs regulate many levels of transcriptional process such as modulating chromatin structure by recruiting coregulators to the transcriptional unit.^{153,154}

During normal mammalian brain development, adolescence seems to be a time of great epigenetic shift. Somel and associates¹⁵⁵ showed that there was heightened level of epigenetic modulation during adolescence, which the authors believed extended neuronal plasticity. Similarly, Lister and colleagues¹⁵⁶ also identified large-scale reconfiguration of the epigenome during periods of heightened synaptogenesis such as during adolescence. These changes in the epigenome likely help to regulate sex differences in the brain. The preoptic area is an area important for sexual, parenting, and thermoregulating behaviors. In this area, the expression of the ER α is modulated by estradiol resulting in 30% greater methylation in females compared with males.¹⁴⁹ In males, higher levels of histone deacetylases play a key role in

programming male sexual behavior.¹⁵⁷ And last, Zinc Fingers, which represses transcription by binding to regulatory regions of DNA, plays a role in modulating pubertal onset in both males and females.^{158,159}

With the elevated level of epigenetic modulation and increased brain flexibility, adolescents are at increased risk of detriment and long-term impairment. In fact, studies were able to show the differential effects of substances on the adolescent brain compared with the adult brain. With repeated exposure to ethanol, adolescent rats showed reduced expression of dopamine receptor D₂, and glutamate ionotropic receptor NMDA type subunit 2 in the prefrontal cortex and altered acetylation levels of histones H3 and H4, which was mediated by elevated histone acetyltransferase (histone deacetylase), in the frontal cortex, striatum, and nucleus accumbens.¹⁶⁰ The same exposure to adult rats showed no changes in the measures as listed. Other studies have shown similar long-term changes to the organization of the reward and emotional circuitry.^{161–164} Nicotine studies showed the greatest gene alterations occurred during the mid-to-late adolescent periods, corresponding with the age of greatest dependence.¹⁶⁵ Similarly, psychostimulant studies showed more pronounced effects in the adolescents compared with adults. Amphetamine and methylphenidate both showed decreased messenger RNA levels of BDNF in the prefrontal cortex and hippocampus but actually increased BDNF levels in adult prefrontal cortex.¹⁶⁶ The attenuation of a factor involved in neuronal growth and plasticity in adolescents is certainly concerning.

But, just as the heightened brain plasticity could result in detrimental effects, an enriched environment could produce benefits. A single week of voluntary exercise in adolescent rats led to significant increases in BDNF, and epigenetic regulators such as histone deacetylase and DNA methyltransferase decreased.¹⁶⁷ With 4 weeks of exercise, the adolescent rats demonstrated improved memory that was sustained into adulthood.¹⁶⁸ Adults who exercised showed immediate benefits in memory enhancement and elevation of BDNF but no long-term benefits. Interestingly, low-to-moderate intensity exercise was sufficient for increased neurogenesis and increased level of BDNF, whereas high-intensity exercise may have triggered a stress response that offset the benefits of exercise.¹⁶⁹ Even more impressive, social enrichment was able to mitigate impact of prenatal alcohol exposure in a rat model of fetal alcohol spectrum disorder.¹⁷⁰

Evolutionarily speaking, adolescence is a time of transition where independent skills are acquired to facilitate the transition into adulthood. Brain plasticity provides opportunity for the developmental changes in TAY, creating a physiologic environment that intersects with the environmental and emotional changes that contribute to independent skill development. Increases in peer-directed social interactions, and intensification of risk taking and novelty seeking behaviors mediated by increases in dopamine levels and enervation may contribute to more successful mating and independence.^{1,171,172} To balance this increase in risk, the increased emotional reactivity may contribute to heightened vigilance of risks and threats thereby increasing opportunity for survival.¹²² Increased epigenetic modulation and brain plasticity may contribute during this time of great change by mediating learning of new information. It was possibly a survival advantage at some point in human evolution, especially when one considers the differences in life expectancy and social pace across the history of human development. However, given the statistics on morbidity and mortality, it is important to consider both the advantages and the potential risks that result from this mismatch in modern TAY. The question as to whether this elevated plasticity is a unique adolescent risk factor or a transition to a more stable brain remains.¹⁷³

SUMMARY

Over the past 2 decades, there have been substantial developments in the understanding of brain development. Progress in neuroimaging has allowed us to better understand the nuances of the development of cortical, subcortical, and white matter structures. Modern neuroscience, genomics, and epigenomic studies allow us a lens through which to develop an understanding of TAY behavior from a neurodevelopmental perspective. We now understand, to a greater degree, why adolescents have difficulty with impulse control, risky behavior, and are disproportionately influenced by their peers. Although our current science has not provided us with all the answers, we can now state confidently that negative environmental influences impact the structure and function of the human brain and the thoughts, actions, and behaviors that result. Armed with emerging evidence that positive environmental influences can influence positively the structure and function of the brain and foment improved academic, memory, mood, and emotional regulatory outcomes,¹⁷⁴ it is now time for child and adolescent psychiatry to design brain-building health promotion and illness prevention approaches for all children, but particularly those in the TAB risk epoch. Developing brain-building health promotion and illness prevention approaches for TAB individuals will likely yield reductions in morbidity and mortality in this high-risk period, enhance individual life trajectories, and possibly have life-long impact on friends and families of TAY.

REFERENCES

- 1. Spear LP. The adolescent brain and age-related behavioral manifestations. Neurosci Biobehav Rev 2000;24(4):417–63.
- Huttenlocher PR, de Courten C, Garey LJ, et al. Synaptogenesis in human visual cortex–evidence for synapse elimination during normal development. Neurosci Lett 1982;33(3):247–52.
- 3. Becker LE, Armstrong DL, Chan F, et al. Dendritic development in human occipital cortical neurons. Brain Res 1984;315(1):117–24.
- Mrzljak L, Uylings HB, Kostovic I, et al. Prenatal development of neurons in the human prefrontal cortex: I. A qualitative Golgi study. J Comp Neurol 1988; 271(3):355–86.
- Mrzljak L, Uylings HB, Kostovic I, et al. Prenatal development of neurons in the human prefrontal cortex. II. A quantitative Golgi study. J Comp Neurol 1992; 316(4):485–96.
- Mrzljak L, Uylings HB, Van Eden CG, et al. Neuronal development in human prefrontal cortex in prenatal and postnatal stages. Prog Brain Res 1990;85: 185–222.
- 7. Bourgeois JP, Goldman-Rakic PS, Rakic P. Synaptogenesis in the prefrontal cortex of rhesus monkeys. Cereb Cortex 1994;4(1):78–96.
- 8. Bourgeois JP. Synaptogenesis, heterochrony and epigenesis in the mammalian neocortex. Acta Paediatr Suppl 1997;422:27–33.
- 9. Rakic P, Bourgeois JP, Eckenhoff MF, et al. Concurrent overproduction of synapses in diverse regions of the primate cerebral cortex. Science 1986;232(4747): 232–5.
- Levitt P. Structural and functional maturation of the developing primate brain. J Pediatr 2003;143(4 Suppl):S35–45.
- 11. Stiles J, Jernigan TL. The basics of brain development. Neuropsychol Rev 2010; 20(4):327–48.

- 12. Bourgeois JP, Rakic P. Changes of synaptic density in the primary visual cortex of the macaque monkey from fetal to adult stage. J Neurosci 1993;13(7): 2801–20.
- 13. Huttenlocher PR, Dabholkar AS. Regional differences in synaptogenesis in human cerebral cortex. J Comp Neurol 1997;387(2):167–78.
- Stanfield BB, O'Leary DD. The transient corticospinal projection from the occipital cortex during the postnatal development of the rat. J Comp Neurol 1985; 238(2):236–48.
- Stanfield BB, O'Leary DD, Fricks C. Selective collateral elimination in early postnatal development restricts cortical distribution of rat pyramidal tract neurones. Nature 1982;298(5872):371–3.
- 16. Innocenti GM, Price DJ. Exuberance in the development of cortical networks. Nat Rev Neurosci 2005;6(12):955–65.
- Isackson PJ, Huntsman MM, Murray KD, et al. BDNF mRNA expression is increased in adult rat forebrain after limbic seizures: temporal patterns of induction distinct from NGF. Neuron 1991;6(6):937–48.
- Huang EJ, Reichardt LF. Neurotrophins: roles in neuronal development and function. Annu Rev Neurosci 2001;24:677–736.
- Bishop KM, Rubenstein JL, O'Leary DD. Distinct actions of Emx1, Emx2, and Pax6 in regulating the specification of areas in the developing neocortex. J Neurosci 2002;22(17):7627–38.
- 20. Nakamura H, Katahira T, Matsunaga E, et al. Isthmus organizer for midbrain and hindbrain development. Brain Res Brain Res Rev 2005;49(2):120–6.
- 21. Kiecker C, Lumsden A. Hedgehog signaling from the ZLI regulates diencephalic regional identity. Nat Neurosci 2004;7(11):1242–9.
- 22. Lumsden A, Keynes R. Segmental patterns of neuronal development in the chick hindbrain. Nature 1989;337(6206):424–8.
- 23. Gavalas A, Ruhrberg C, Livet J, et al. Neuronal defects in the hindbrain of Hoxa1, Hoxb1 and Hoxb2 mutants reflect regulatory interactions among these Hox genes. Development 2003;130(23):5663–79.
- 24. Stiles J. The fundamentals of brain development: integrating nature and nurture. Cambridge (MA): Harvard University Press; 2008.
- 25. Bishop KM, Goudreau G, O'Leary DD. Regulation of area identity in the mammalian neocortex by Emx2 and Pax6. Science 2000;288(5464):344–9.
- Hamasaki T, Leingärtner A, Ringstedt T, et al. EMX2 regulates sizes and positioning of the primary sensory and motor areas in neocortex by direct specification of cortical progenitors. Neuron 2004;43:359–72.
- 27. Greenough WT, Black JE, Wallace CS. Experience and brain development. Child Dev 1987;58(3):539–59.
- 28. Hubel DH, Wiesel TN. Ferrier lecture. Functional architecture of macaque monkey visual cortex. Proc R Soc Lond B Biol Sci 1977;198(1130):1–59.
- 29. Hubel DH, Wiesel TN, LeVay S. Plasticity of ocular dominance columns in monkey striate cortex. Philos Trans R Soc Lond B Biol Sci 1977;278(961):377–409.
- **30.** Bachevalier J, Mishkin M. Effects of selective neonatal temporal lobe lesions on visual recognition memory in rhesus monkeys. J Neurosci 1994;14(4):2128–39.
- **31.** Hanson JL, Hair N, Shen DG, et al. Family poverty affects the rate of human infant brain growth. PLoS ONE 2013;8(12):e80954.
- Hanson JL, Nacewicz BM, Sutterer MJ, et al. Behavioral problems after early life stress: contributions of the hippocampus and amygdala. Biol Psychiatry 2015; 77(4):314–23.

- **33.** Luby JL, Belden AC, Jackson JJ, et al. Early childhood depression and alterations in the trajectory of gray matter maturation in middle childhood and early adolescence. JAMA Psychiatry 2016;73(1):31–8.
- 34. Belden AC, Barch DM, Oakberg TJ, et al. Anterior insula volume and guilt: neurobehavioral markers of recurrence after early childhood major depressive disorder. JAMA Psychiatry 2015;72(1):40–8.
- **35.** Luby J, Lenze S, Tillman R. A novel early intervention for preschool depression: findings from a pilot randomized controlled trial. J Child Psychol Psychiatry 2012;53(3):313–22.
- **36.** Burghy CA, Stodola DE, Ruttle PL, et al. Developmental pathways to amygdalaprefrontal function and internalizing symptoms in adolescence. Nat Neurosci 2012;15(12):1736–41.
- Fan Y, Herrera-Melendez AL, Pestke K, et al. Early life stress modulates amygdala-prefrontal functional connectivity: implications for oxytocin effects. Hum Brain Mapp 2014;35(10):5328–39.
- Gee DG, Gabard-Durnam LJ, Flannery J, et al. Early developmental emergence of human amygdala-prefrontal connectivity after maternal deprivation. Proc Natl Acad Sci USA 2013;110(39):15638–43.
- Grant MM, Wood K, Sreenivasan K, et al. Influence of early life stress on intraand extra-amygdaloid causal connectivity. Neuropsychopharmacology 2015; 40(7):1782–93.
- **40.** Barch D, Pagliaccio D, Belden A, et al. Effect of hippocampal and amygdala connectivity on the relationship between preschool poverty and school-age depression. Am J Psychiatry 2016;173(6):625–34.
- 41. Hebb DO. The organization of behavior. New York: Wiley; 1949.
- 42. Diamond MC, Krech D, Rosenzweig MR. The effects of an enriched environment on the histology of the rat cerebral cortex. J Comp Neurol 1964;123:111–20.
- Rosenzweig MR, Bennett EL, Diamond MC. Brain changes in response to experience. Scientific American. 1972. Available at: http://psycnet.apa.org/psycinfo/ 1972-22480-001. Accessed October 4, 2016.
- 44. Bennett EL, Rosenzweig MR. Difference in occipital cortical synapses from environmentally enriched, impoverished, and standard colony rats. J Neurosci Res 1975;1(2):109–19.
- 45. Bennett EL, Rosenzweig MR. Quantitative synaptic changes with differential experience in rat brain. Int J Neurosci 1971;2(3):113–27.
- 46. Bennett EL, Rosenzweig MR, Diamond MC. Rat brain: effects of environmental enrichment on wet and dry weights. Science 1969;163(3869):825–6. Available at: http://science.sciencemag.org/content/163/3869/825.short.
- Rosenzweig MR, Bennett EL, Diamond MC. Effects of differential environments on brain anatomy and brain chemistry. Proc Annu Meet Am Psychopathol Assoc 1967;56:45–56.
- Diamond MC, Rosenzweig MR, Bennett EL, et al. Effects of environmental enrichment and impoverishment on rat cerebral cortex. J Neurobiol 1972;3(1): 47–64.
- Diamond MC, Law F, Rhodes H, et al. Increases in cortical depth and glia numbers in rats subjected to enriched environment. J Comp Neurol 1966; 128(1):117–26.
- Rosenzweig MR, Bennett EL, Diamond MC, et al. Influences of environmental complexity and visual stimulation on development of occipital cortex in rat. Brain Res 1969;14(2):427–45.

- 51. Black JE, Sirevaag AM, Greenough WT. Complex experience promotes capillary formation in young rat visual cortex. Neurosci Lett 1987;83(3):351–5.
- Greenough WT, Chang FL. Dendritic pattern formation involves both oriented regression and oriented growth in the barrels of mouse somatosensory cortex. Brain Res 1988;471(1):148–52.
- Jones TA, Greenough WT. Ultrastructural evidence for increased contact between astrocytes and synapses in rats reared in a complex environment. Neurobiol Learn Mem 1996;65(1):48–56.
- 54. Markham JA, Greenough WT. Experience-driven brain plasticity: beyond the synapse. Neuron Glia Biol 2004;1(4):351–63.
- 55. Chang FL, Greenough WT. Lateralized effects of monocular training on dendritic branching in adult split-brain rats. Brain Res 1982;232(2):283–92.
- 56. Sirevaag AM, Greenough WT. Differential rearing effects on rat visual cortex synapses. III. Neuronal and glial nuclei, boutons, dendrites, and capillaries. Brain Res 1987;424(2):320–32.
- 57. Uylings HB, Kuypers K, Veltman WA. Environmental influences on the neocortex in later life. Prog Brain Res 1978;48:261–74.
- Uylings HB, Kuypers K, Diamond MC, et al. Effects of differential environments on plasticity of dendrites of cortical pyramidal neurons in adult rats. Exp Neurol 1978;62(3):658–77.
- **59.** Boyke J, Driemeyer J, Gaser C, et al. Training-induced brain structure changes in the elderly. J Neurosci 2008;28(28):7031–5.
- 60. Driemeyer J, Boyke J, Gaser C, et al. Changes in gray matter induced by learning-revisited. PLoS ONE 2008;3(7):e2669.
- **61.** Draganski B, Gaser C, Busch V, et al. Neuroplasticity: changes in grey matter induced by training. Nature 2004;427(6972):311–2.
- 62. Pantev C, Okamoto H, Teismann H. Music-induced cortical plasticity and lateral inhibition in the human auditory cortex as foundations for tonal tinnitus treatment. Front Syst Neurosci 2012;6:50.
- **63.** Pantev C, Roberts LE, Schulz M, et al. Timbre-specific enhancement of auditory cortical representations in musicians. Neuroreport 2001;12(1):169–74.
- 64. Pantev C, Oostenveld R, Engelien A, et al. Increased auditory cortical representation in musicians. Nature 1998;392(6678):811–4.
- Hudziak JJ, Albaugh MD, Ducharme S, et al. Cortical thickness maturation and duration of music training: health-promoting activities shape brain development. J Am Acad Child Adolesc Psychiatry 2014;53(11):1153–61, 1161.e1–2.
- **66.** Bengtsson SL, Nagy Z, Skare S, et al. Extensive piano practicing has regionally specific effects on white matter development. Nat Neurosci 2005;8(9):1148–50.
- 67. Scholz J, Klein MC, Behrens TE, et al. Training induces changes in white-matter architecture. Nat Neurosci 2009;12(11):1370–1.
- **68.** Takeuchi H, Taki Y, Nouchi R, et al. Working memory training impacts the mean diffusivity in the dopaminergic system. Brain Struct Funct 2015;220(6):3101–11.
- 69. Takeuchi H, Sekiguchi A, Taki Y, et al. Training of working memory impacts structural connectivity. J Neurosci 2010;30(9):3297–303.
- **70.** Jernigan TL, Tallal P. Late childhood changes in brain morphology observable with MRI. Dev Med Child Neurol 1990;32(5):379–85.
- 71. Jernigan TL, Trauner DA, Hesselink JR, et al. Maturation of human cerebrum observed in vivo during adolescence. Brain 1991;114(Pt 5):2037–49.
- 72. Giedd JN, Blumenthal J, Jeffries NO, et al. Brain development during childhood and adolescence: a longitudinal MRI study. Nat Neurosci 1999;2(10):861–3.

- 73. Giedd JN, Snell JW, Lange N, et al. Quantitative magnetic resonance imaging of human brain development: ages 4-18. Cereb Cortex 1996;6(4):551–60.
- 74. Shaw P, Kabani NJ, Lerch JP, et al. Neurodevelopmental trajectories of the human cerebral cortex. J Neurosci 2008;28(14):3586–94.
- 75. Raznahan A, Shaw P, Lalonde F, et al. How does your cortex grow? J Neurosci 2011;31(19):7174–7.
- **76.** Sowell ER, Thompson PM, Leonard CM, et al. Longitudinal mapping of cortical thickness and brain growth in normal children. J Neurosci 2004;24(38):8223–31.
- 77. Muftuler LT, Davis EP, Buss C, et al. Cortical and subcortical changes in typically developing preadolescent children. Brain Res 2011;1399:15–24.
- **78.** Ducharme S, Albaugh MD, Nguyen T-VV, et al. Trajectories of cortical thickness maturation in normal brain development–The importance of quality control procedures. Neuroimage 2016;125:267–79.
- **79.** Koolschijn PC, Crone EA. Sex differences and structural brain maturation from childhood to early adulthood. Dev Cogn Neurosci 2013;5:106–18.
- **80.** Mutlu AK, Schneider M, Debbané M, et al. Sex differences in thickness, and folding developments throughout the cortex. Neuroimage 2013;82:200–7.
- Zatorre RJ, Fields RD, Johansen-Berg H. Plasticity in gray and white: neuroimaging changes in brain structure during learning. Nat Neurosci 2012;15(4): 528–36.
- 82. Fischl B, Dale AM. Measuring the thickness of the human cerebral cortex from magnetic resonance images. Proc Natl Acad Sci USA 2000;97(20):11050–5.
- Kabani N, Goualher Le G, MacDonald D, et al. Measurement of cortical thickness using an automated 3-D algorithm: a validation study. Neuroimage 2001; 13(2):375–80.
- Gogtay N, Giedd JN, Lusk L, et al. Dynamic mapping of human cortical development during childhood through early adulthood. Proc Natl Acad Sci USA 2004;101(21):8174–9.
- Sowell ER, Thompson PM, Holmes CJ, et al. In vivo evidence for postadolescent brain maturation in frontal and striatal regions. Nat Neurosci 1999; 2(10):859–61.
- **86.** Giedd JN, Rapoport JL. Structural MRI of pediatric brain development: what have we learned and where are we going? Neuron 2010;67(5):728–34.
- 87. Olson IR, Plotzker A, Ezzyat Y. The Enigmatic temporal pole: a review of findings on social and emotional processing. Brain 2007;130(Pt 7):1718–31.
- 88. Shaw P, Greenstein D, Lerch J, et al. Intellectual ability and cortical development in children and adolescents. Nature 2006;440(7084):676–9.
- 89. Casey BJ, Trainor R, Giedd J, et al. The role of the anterior cingulate in automatic and controlled processes: a developmental neuroanatomical study. Dev Psy-chobiol 1997;30(1):61–9.
- **90.** Lu L, Leonard C, Thompson P, et al. Normal developmental changes in inferior frontal gray matter are associated with improvement in phonological processing: a longitudinal MRI analysis. Cereb Cortex 2007;17(5):1092–9.
- **91.** Sowell ER, Thompson PM, Toga AW. Mapping changes in the human cortex throughout the span of life. Neuroscientist 2004;10(4):372–92.
- **92.** Nguyen T-VV, McCracken JT, Albaugh MD, et al. A testosterone-related structural brain phenotype predicts aggressive behavior from childhood to adulthood. Psychoneuroendocrinology 2016;63:109–18.
- Ducharme S, Albaugh MD, Hudziak JJ, et al. Anxious/depressed symptoms are linked to right ventromedial prefrontal cortical thickness maturation in healthy children and young adults. Cereb Cortex 2014;24(11):2941–50.

- **94.** Albaugh MD, Ducharme S, Collins DL, et al. Evidence for a cerebral cortical thickness network anti-correlated with amygdalar volume in healthy youths: implications for the neural substrates of emotion regulation. Neuroimage 2013; 71:42–9.
- **95.** Ducharme S, Hudziak JJ, Botteron KN, et al. Decreased regional cortical thickness and thinning rate are associated with inattention symptoms in healthy children. J Am Acad Child Adolesc Psychiatry 2012;51(1):18–27.e2.
- **96.** Ameis SH, Ducharme S, Albaugh MD, et al. Cortical thickness, corticoamygdalar networks, and externalizing behaviors in healthy children. Biol Psychiatry 2014;75(1):65–72.
- **97.** Giedd JN, Vaituzis AC, Hamburger SD, et al. Quantitative MRI of the temporal lobe, amygdala, and hippocampus in normal human development: ages 4-18 years. J Comp Neurol 1996;366(2):223–30.
- **98.** Toga AW, Thompson PM, Sowell ER. Mapping brain maturation. Trends Neurosci 2006;29(3):148–59.
- **99.** Sowell ER, Trauner DA, Gamst A, et al. Development of cortical and subcortical brain structures in childhood and adolescence: a structural MRI study. Dev Med Child Neurol 2002;44(1):4–16.
- 100. Ostby Y, Tamnes CK, Fjell AM, et al. Heterogeneity in subcortical brain development: a structural magnetic resonance imaging study of brain maturation from 8 to 30 years. J Neurosci 2009;29(38):11772–82.
- 101. Giedd JN, Clasen LS, Lenroot R, et al. Puberty-related influences on brain development. Mol Cell Endocrinol 2006;254-255:154–62.
- 102. Lenroot RK, Giedd JN. Brain development in children and adolescents: insights from anatomical magnetic resonance imaging. Neurosci Biobehav Rev 2006; 30(6):718–29.
- 103. Guo X, Chen C, Chen K, et al. Brain development in Chinese children and adolescents: a structural MRI study. Neuroreport 2007;18(9):875–80.
- 104. Gogtay N, Nugent TF, Herman DH, et al. Dynamic mapping of normal human hippocampal development. Hippocampus 2006;16(8):664–72.
- **105.** Benes FM, Turtle M, Khan Y, et al. Myelination of a key relay zone in the hippocampal formation occurs in the human brain during childhood, adolescence, and adulthood. Arch Gen Psychiatry 1994;51(6):477–84.
- 106. Paus T, Collins DL, Evans AC, et al. Maturation of white matter in the human brain: a review of magnetic resonance studies. Brain Res Bull 2001;54(3): 255–66.
- 107. Walhovd KB, Fjell AM, Reinvang I, et al. Effects of age on volumes of cortex, white matter and subcortical structures. Neurobiol Aging 2005;26(9):1261–70 [discussion: 1275–8].
- 108. Hermoye L, Saint-Martin C, Cosnard G, et al. Pediatric diffusion tensor imaging: normal database and observation of the white matter maturation in early child-hood. Neuroimage 2006;29(2):493–504.
- 109. Li TQ, Noseworthy MD. Mapping the development of white matter tracts with diffusion tensor imaging. Developmental Science 2002;5:293–300.
- 110. Imperati D, Colcombe S, Kelly C, et al. Differential development of human brain white matter tracts. PLoS ONE 2011;6(8):e23437.
- 111. Luna B, Garver KE, Urban TA, et al. Maturation of cognitive processes from late childhood to adulthood. Child Dev 2004;75(5):1357–72.
- 112. Luna T, Munoz M. Maturation of widely distributed brain function subserves cognitive development. Neuroimage 2001;13(5):786–93.

- 113. Zald I. The development of spatial working memory abilities. Dev Neuropsychol 2004;26(1):487–512.
- 114. Dahl RE. Adolescent brain development: a period of vulnerabilities and opportunities. Keynote address. Ann N Y Acad Sci 2004;1021:1–22.
- 115. Dahl RE, Gunnar MR. Heightened stress responsiveness and emotional reactivity during pubertal maturation: implications for psychopathology. Dev Psychopathol 2009;21(1):1–6.
- 116. Kessler BP, Berglund P, Demler O, et al. Lifetime prevalence and age-of-onset distribution of DSM-IV disorders in the National Comorbidity Survey Replication. Arch Gen Psychiatry 2005;62:593–603. Available at: https://www.ncbi.nlm.nih.gov/ pubmed/?term=Demler%200%5BAuthor%5D&cauthor=true&cauthor_uid=159 39837.
- 117. Giedd JN. The amazing teen brain. Sci Am 2015;312(6):32–7.
- **118.** Gabard-Durnam LJ, Flannery J, Goff B, et al. The development of human amygdala functional connectivity at rest from 4 to 23 years: a cross-sectional study. Neuroimage 2014;95:193–207.
- 119. Yurgelun-Todd D. Emotional and cognitive changes during adolescence. Curr Opin Neurobiol 2007;17(2):251–7.
- 120. Dahl RE. Biological, developmental, and neurobehavioral factors relevant to adolescent driving risks. Am J Prev Med 2008;35(3 Suppl):S278–84.
- 121. Mills KL, Goddings A-LL, Clasen LS, et al. The developmental mismatch in structural brain maturation during adolescence. Dev Neurosci 2014;36(3–4): 147–60.
- 122. Casey BJ, Jones RM, Hare TA. The adolescent brain. Ann N Y Acad Sci 2008; 1124:111–26.
- 123. Liston C, McEwen BS, Casey BJ. Psychosocial stress reversibly disrupts prefrontal processing and attentional control. Proc Natl Acad Sci USA 2009; 106(3):912–7.
- 124. Rubia K, Halari R, Smith AB, et al. Dissociated functional brain abnormalities of inhibition in boys with pure conduct disorder and in boys with pure attention deficit hyperactivity disorder. Am J Psychiatry 2008;165(7):889–97.
- 125. Stevens MC, Kiehl KA, Pearlson GD, et al. Functional neural networks underlying response inhibition in adolescents and adults. Behav Brain Res 2007;181(1): 12–22.
- 126. Andersen SL. Changes in the second messenger cyclic AMP during development may underlie motoric symptoms in attention deficit/hyperactivity disorder (ADHD). Behav Brain Res 2002;130(1–2):197–201.
- 127. Galvan A, Hare TA, Parra CE, et al. Earlier development of the accumbens relative to orbitofrontal cortex might underlie risk-taking behavior in adolescents. J Neurosci 2006;26(25):6885–92.
- 128. Cohen JR, Asarnow RF, Sabb FW, et al. A unique adolescent response to reward prediction errors. Nat Neurosci 2010;13(6):669–71.
- 129. Van Leijenhorst L, Gunther Moor B, Op de Macks ZA, et al. Adolescent risky decision-making: neurocognitive development of reward and control regions. Neuroimage 2010;51(1):345–55.
- Schneider S, Peters J, Bromberg U, et al. Risk taking and the adolescent reward system: a potential common link to substance abuse. Am J Psychiatry 2012; 169(1):39–46.
- 131. Ernst M, Nelson EE, Jazbec S, et al. Amygdala and nucleus accumbens in responses to receipt and omission of gains in adults and adolescents. Neuroimage 2005;25(4):1279–91.

- 132. Bjork JM, Smith AR, Danube CL, et al. Developmental differences in posterior mesofrontal cortex recruitment by risky rewards. J Neurosci 2007;27(18): 4839–49.
- 133. Steinberg LA. Social neuroscience perspective on adolescent risk-taking. Dev Rev 2008;28(1):78–106.
- **134.** Steinberg L, Graham S, O'Brien L, et al. Age differences in future orientation and delay discounting. Child Dev 2009;80(1):28–44.
- 135. Hare TA, Tottenham N, Galvan A, et al. Biological substrates of emotional reactivity and regulation in adolescence during an emotional go-nogo task. Biol Psychiatry 2008;63(10):927–34.
- **136.** Klingler J, Gloor P. The connections of the amygdala and of the anterior temporal cortex in the human brain. J Comp Neurol 1960;115:333–69.
- 137. Ebeling U, von Cramon D. Topography of the uncinate fascicle and adjacent temporal fiber tracts. Acta Neurochir (Wien) 1992;115(3–4):143–8.
- **138.** Lebel C, Walker L, Leemans A, et al. Microstructural maturation of the human brain from childhood to adulthood. Neuroimage 2008;40(3):1044–55.
- 139. Banks SJ, Eddy KT, Angstadt M, et al. Amygdala-frontal connectivity during emotion regulation. Soc Cogn Affect Neurosci 2007;2(4):303–12.
- 140. Phillips ML, Drevets WC, Rauch SL, et al. Neurobiology of emotion perception I: the neural basis of normal emotion perception. Biol Psychiatry 2003;54(5): 504–14.
- 141. Hanson JL, Chung MK, Avants BB, et al. Early stress is associated with alterations in the orbitofrontal cortex: a tensor-based morphometry investigation of brain structure and behavioral risk. J Neurosci 2010;30:7466–72.
- 142. Lupien SJ, Parent S, Evans AC, et al. Larger amygdala but no change in hippocampal volume in 10-year-old children exposed to maternal depressive symptomatology since birth. Proc Natl Acad Sci U S A 2011;108:14324–9.
- 143. Nacewicz BM, Dalton KM, Johnstone T, et al. Amygdala volume and nonverbal social impairment in adolescent and adult males with autism. Arch Gen Psychiatry 2006;63:1417–28.
- 144. Mosconi MW, Cody-Hazlett H, Poe MD, et al. Longitudinal study of amygdala volume and joint attention in 2- to 4-year-old children with autism. Arch Gen Psychiatry 2009;66:509–16.
- 145. Davidson RJ, McEwen BS. Social influences on neuroplasticity: stress and interventions to promote well-being. Nat Neurosci 2012;15(5):689–95.
- 146. Sheline YI. 3D MRI studies of neuroanatomic changes in unipolar major depression: the role of stress and medical comorbidity. Biol Psychiatry 2000;48(8): 791–800.
- 147. Kasai K, Yamasue H, Gilbertson MW, et al. Evidence for acquired pregenual anterior cingulate gray matter loss from a twin study of combat-related posttraumatic stress disorder. Biol Psychiatry 2008;63(6):550–6.
- 148. Champagne FA. Epigenetic influence of social experiences across the lifespan. Dev Psychobiol 2010;52(4):299–311.
- 149. McCarthy MM, Nugent BM. Epigenetic contributions to hormonally-mediated sexual differentiation of the brain. J Neuroendocrinol 2013;25(11):1133–40.
- 150. Chahrour M, Jung SY, Shaw C, et al. MeCP2, a key contributor to neurological disease, activates and represses transcription. Science 2008;320(5880): 1224–9.
- 151. Fagiolini M, Jensen CL, Champagne FA. Epigenetic influences on brain development and plasticity. Curr Opin Neurobiol 2009;19(2):207–12.

- 152. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 2004;116(2):281–97.
- 153. Mattick JS, Amaral PP, Dinger ME, et al. RNA regulation of epigenetic processes. Bioessays 2009;31(1):51–9.
- 154. Mehler MF. Epigenetic principles and mechanisms underlying nervous system functions in health and disease. Prog Neurobiol 2008;86(4):305–41.
- 155. Somel M, Franz H, Yan Z, et al. Transcriptional neoteny in the human brain. Proc Natl Acad Sci USA 2009;106(14):5743–8.
- **156.** Lister R, Mukamel EA, Nery JR, et al. Global epigenomic reconfiguration during mammalian brain development. Science 2013;341(6146):1237905.
- 157. Matsuda KI, Mori H, Nugent BM, et al. Histone deacetylation during brain development is essential for permanent masculinization of sexual behavior. Endocrinology 2011;152(7):2760–7.
- 158. Lomniczi A, Wright H, Castellano JM, et al. Epigenetic regulation of puberty via Zinc finger protein-mediated transcriptional repression. Nat Commun 2015;6: 10195.
- 159. Lomniczi A, Loche A, Castellano JM, et al. Epigenetic control of female puberty. Nat Neurosci 2013;16(3):281–9.
- 160. Pascual M, Boix J, Felipo V, et al. Repeated alcohol administration during adolescence causes changes in the mesolimbic dopaminergic and glutamatergic systems and promotes alcohol intake in the adult rat. J Neurochem 2009; 108(4):920–31.
- **161.** Coleman LG, He J, Lee J, et al. Adolescent binge drinking alters adult brain neurotransmitter gene expression, behavior, brain regional volumes, and neuro-chemistry in mice. Alcohol Clin Exp Res 2011;35(4):671–88.
- **162.** Pandey SC, Sakharkar AJ, Tang L, et al. Potential role of adolescent alcohol exposure-induced amygdaloid histone modifications in anxiety and alcohol intake during adulthood. Neurobiol Dis 2015;82:607–19.
- **163.** Kyzar EJ, Zhang H, Sakharkar AJ, et al. Adolescent alcohol exposure alters lysine demethylase 1 (LSD1) expression and histone methylation in the amygdala during adulthood. Addict Biol 2016. [Epub ahead of print].
- **164.** Sakharkar AJ, Tang L, Zhang H, et al. Effects of acute ethanol exposure on anxiety measures and epigenetic modifiers in the extended amygdala of adolescent rats. Int J Neuropsychopharmacol 2014;17(12):2057–67.
- 165. Polesskaya OO, Fryxell KJ, Merchant AD, et al. Nicotine causes age-dependent changes in gene expression in the adolescent female rat brain. Neurotoxicol Teratol 2007;29(1):126–40.
- 166. Banerjee PS, Aston J, Khundakar AA, et al. Differential regulation of psychostimulant-induced gene expression of brain derived neurotrophic factor and the immediate-early gene Arc in the juvenile and adult brain. Eur J Neurosci 2009;29(3):465–76.
- 167. Abel JL, Rissman EF. Running-induced epigenetic and gene expression changes in the adolescent brain. Int J Dev Neurosci 2013;31(6):382–90.
- **168.** Hopkins ME, Nitecki R, Bucci DJ. Physical exercise during adolescence versus adulthood: differential effects on object recognition memory and brain-derived neurotrophic factor levels. Neuroscience 2011;194:84–94.
- 169. Lou SJ, Liu JY, Chang H, et al. Hippocampal neurogenesis and gene expression depend on exercise intensity in juvenile rats. Brain Res 2008;1210:48–55.
- 170. Ignacio C, Mooney SM, Middleton FA. Effects of acute prenatal exposure to ethanol on microRNA expression are ameliorated by social enrichment. Front Pediatr 2014;2:103.

- 171. Rosenberg DR, Lewis DA. Postnatal maturation of the dopaminergic innervation of monkey prefrontal and motor cortices: a tyrosine hydroxylase immunohisto-chemical analysis. J Comp Neurol 1995;358(3):383–400.
- 172. Laviola G, Adriani W, Terranova ML, et al. Psychobiological risk factors for vulnerability to psychostimulants in human adolescents and animal models. Neurosci Biobehav Rev 1999;23(7):993–1010.
- 173. Spear LP. Adolescent neurodevelopment. J Adolesc Health 2013;52(2 Suppl 2): S7–13.
- 174. O'Loughlin K, Hudziak JJ. Health promotion and prevention in child and adolescent mental health. In: Rey JM, editor. IACAPAP Textbook of Child and Adolescent Mental Health. Geneva (Switzerland): International Association for Child and Adolescent Psychiatry and Allied Professions; 2017. p. 1–23.